



# **NEw Technologies for macro and microplastic Detection and**Analysis in the Adriatic Basin

**NET4mPlastic** 

# **ADRI**atic Sea and **PLASTi**c and microplastic pollution from freshwaters – an Adrion territorial challenge

**AdriPlast** 

PhD Corinne Corbau – cbc@unife.it

Conference on Microplastics in drinking water – from source to tap EU Strategy for the Danube Region Water Quality Priority Area

# NET4mPLAST GENERAL INFORMATION

2014 - 2020 Interreg V-A Italy - Croatia CBC Programme

Call for proposal 2017 Standard

Priority Axis: Environment and cultural heritage

Specific Objective 3.3 - Improve the environmental quality conditions of the sea and coastal area by use of sustainable and innovative technologies and approaches

€ 2.478.640,00

Start date: 01/01/2019

End: 30/06/2022

### **Partnership**

- University of Ferrara, IT (LP)
- University of Trieste, IT
- Marche Region, IT
- HYDRA SOLUTIONS SRL, IT
- PROSOFT LTD, HR
- Istituto Zooprofilatico Sperimentale dell'Abruzzo e del Molise 'G. Caporale', IT
- Teaching Institute for Public Health,
   Primorje-Gorski Kotar County, HR
- Public Institution RERA, HR
- University of Split, HR





















#### **Programme output indicator**

3.301 Environmental friendly technological solutions (and approaches) implemented

#### **Project main outputs**

- Open access common platform coupled with an Early Warning System allowing the sharing of real time and forecast data and the planning of common actions
- On-board unit for field activities and installed on marine drones for activities along the coast and on ships for offshore activities

## **Programme output indicator**

3.303 Microplastic waste collected in marine areas

#### **Project main outputs**

- Definition of a Joint methodology for MP sampling, monitoring and analysis

#### **Target groups**

- Local, regional and national public authorities (municipalities for instance)
- Regional and local development agencies
- Associations, innovation and environmental agencies
- Education and training organizations as well as universities and research institutes



## **ADRIPLAST GENERAL INFORMATION**

**Interreg IPA Adrion 2021-2027** 

First call for proposal 2023

Programme priority: Supporting a greener and climate resilient Adriatic - Ionian region

SO 2.3: Supporting environment preservation and protection in the Adriatic - Ionian region

LP: CURSA

€ 1.428.477,65

Start date: 01/09/2024

36 months

EU part of cooperation area

Non-EU part of cooperation area

Just started Open for collaboration

### **Partnership**

- CURSA (LP, IT)
- University of Ferrara Italia (IT)
- Ruđer Bošković Institute Hrvatska (HR)
- National institute of biology Slovenija (SI)
- Inst. for Biological Research "Siniša Stanković" - National Institute of the Rep. of Serbia (RS)
- Institute of Public Health of Montenegro Crna (ME)
- National Center of Environmental Movement Shqipëria (AL)

#### **Associated Partner:**

Istituto Zooprofilattico Sperimentale dell'Abruzzo e del

Molise "G. Caporale" (IZS Teramo)























# ADRIPLAST GENERAL INFORMATION

### Main objectives

Enhancing transnational collaboration Identifying and addressing key challenges Developing sustainable solutions.

### **Key Outputs**

- Enhanced comprehension of plastic pollution dimensions within pilot sites
- A roadmap for monitoring and mitigating plastic
- Transnational networks and knowledge sharing
- Awareness-raising and capacity-building activities

Work package 1: Preliminary activities and initial state

Work package 2: Plastic and microplastic pollution (PmPP) from source to sea

Work package 3: Waste managment and possible solutions

## **Target groups**

Local public authority - Regional public authority - Interest groups including NGOs - Higher education and research organisations - General public - Cross-border legal body - Education/training center and school

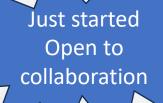
EU part of cooperation area

Non-EU part of cooperation area









#### **Preliminary activities**

#### Data collection for selected Macro-Areas:

- Morphological, sedimentological, and hydrodynamic characteristics
- MP occurrence (beach, sea surface, sea bottom and biota) from literature and EU projects' results
- Characteristics of other relevant water parameters (transparency, temperature, salinity, dissolved oxygen (% and mg/l), pH, depth, flow, etc.)
- Meteo-marine data record

Application of a 3D transport model for MP accumulation/deposition areas

Development of the **Integrated Platform's** structure for data analysis and sharing

#### **Field activities**

Water, sediment and biota sampling (traditional /innovative) for laboratory analysis in areas of MP accumulation and MP nonaccumulation

**Real-time analysis** of parameters relevant to MP (transparency, temperature, salinity, dissolved oxygen (% and mg/l), pH, depth, localization, flow, etc.)

## **Laboratory activities**

Cost-effective lab. analysis procedures for reducing time consumption:

- MP content in all the marine compartments
- Occurrence of related co-contaminant sorbed by MP
- other relevant parameters

Analysis of time of permanence of MP and co-contaminant in sampled biota

Analysis of human health risks

Implementation of the **Integrated platform** 

Setting and calibration of the **Early-Warning System** 

#### **Outputs:**

- Validated model for MP accumulation areas prediction
- Potential scenarios forecasting of MP accumulation areas (MP-vulnerability map creation)
- Early Warning System for MP accumulation areas forecasted after storms/river overflows
- Indication to aquaculture and fishing operators, local authorities, stakeholders for safety food production
- Indication to local authorities for MP vulnerability mitigation
- Shared GIS database
- Cost-effective procedures standardisation
- Formation of operators and technicians



## Demonstration activities: test sites

## Monitoring plastic and microplastic wastes on coastal and marine environments

In situ surveys have been carried out twice/year and before and after an extreme event (i.e. flooding or sea-storm).

These activities provided a large pool of samples regarding plastic contamination.



The surveys have been performed adapting the **De FishGear protocols\*** 



#### **Beach Sampling:**

- area identification (100 m perpendicular to the coastline)
- subdivision into 10 transepts (~ every 10 m)
- Marine Litter sampling (2,5cm < x < 50cm)</li>
- LMP sampling (mesoplastic) (<2,5cm) on 1mq: 3l within the first 3 cm (x6)</li>
- SMP sampling (microplastics) (0.1 μm <x <5 mm): 250ml (x6)</li>

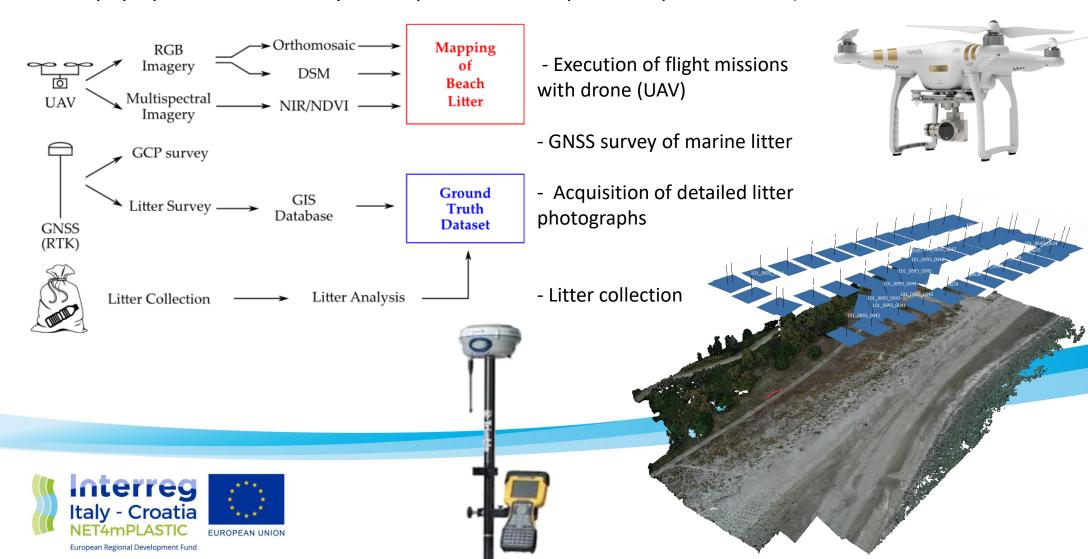




European Regional Development Fund

## **Monitoring of marine litter**

- Field surveys (using innovative and traditional approach) for morphology monitoring and plastic detection:
  - **GPS** (topography and bathymetry)
  - UAV based monitoring system and remote sensing (multispectral data, ocean colour, phytoplankton, chemistry, atmospheric water vapour, temperature, etc.)



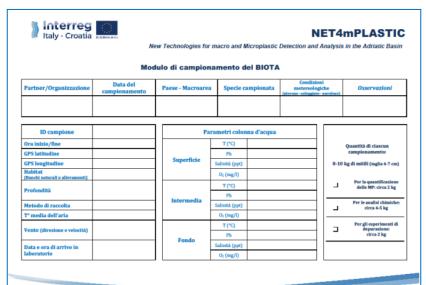
## Monitoring marine litter in coastal waters

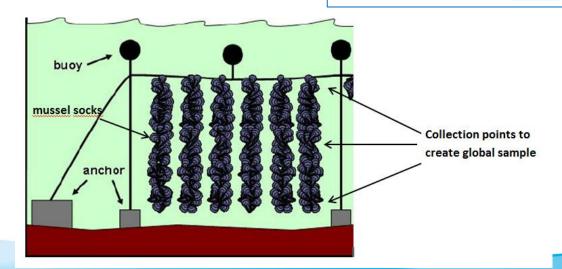
- Sea Surface Sampling: area identif
- area identification (sampling beach area-facing)
  - 4 transects manta trawl towed by boat along the transepts for ~1/2 hour each
  - collection of trapped material and storage in glass jars with alcoholic solution

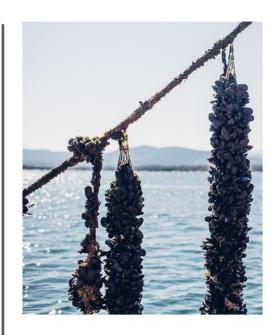


## **Monitoring in Biota - mussels**

- commercial size: 4-7 cm
- Global sample consisting of 3 elementary samples that will be carried out at three depth levels:
  - greater depth
  - in the middle
  - at 50 cm from the surface







- Aluminum foil
- Balance
- Blotting paper
- Caliper
- Latex gloves without powder
- Multi-parameter probe
- Portable refrigerator
- Scissors
- Scalpels
- Water resistant pens



## Lab's analysis

#### **Macro Litter**

2.5 - 50 cm

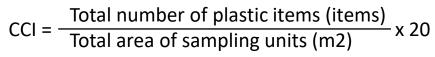




polymer	synthetic rop e, fishing net, other plastic/polystyrene items(identifiable)
materials	
Rubber	<ul> <li>balloons and balloon sticks, rubber boots, tires and belts, bobbins (fishing), condoms (incl packaging), other rubber pieces</li> </ul>
1	
Cloth/	<ul> <li>clothing, shoes, bags, carpets, furnishing, ropes, string and nets etc.</li> </ul>
Textile	
Paper/Ca	cardboard (boxes & fragments), cigarette packets, drink containers, newspapers etc.
rdboard	
Processed /worked	• pallets, corks, processed wood etc.
Metal	<ul> <li>cans (food, aerosol, drinks), refrigerators, washers, fishing related, car parts, batteries, cables, Large metallic objects, etc.</li> </ul>
\ /	A1 A1
Glass/ ceramics	<ul> <li>bottles incl. pieces, jars incl. pieces, glass or ceramic fragments &gt;2.5cm, large glass objects (specify), other glass items.</li> </ul>
-	

oags, bottles, food containers incl. fast food containers, crates and ontainers, plastic caps and lids, cigaretre butts and filters, gloves



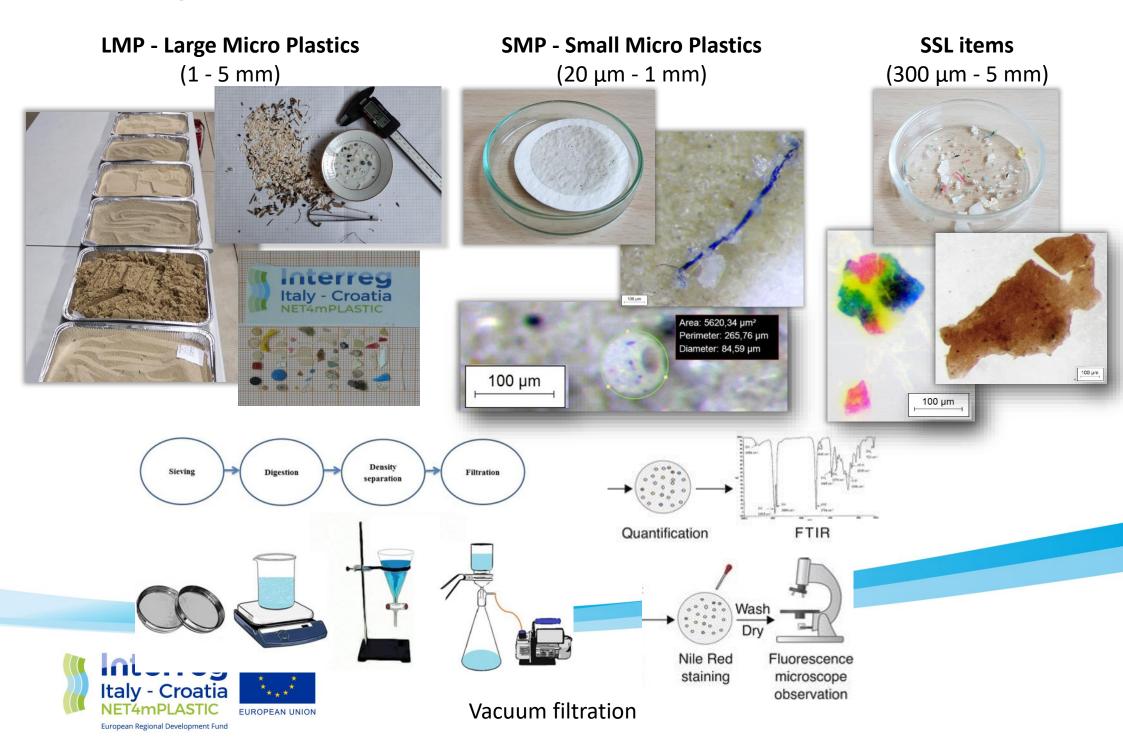


CLEAN COAST INDEX

CLEAN COAST INDEA			
Values	Cleanliness		
0 – 2	Very clean		
0-2	(no litter is seen)		
	Clean		
2 - 5	(no litter is seen over a large		
	area)		
	Moderate		
5 – 10	(a few pieces of litter can be		
	detected)		
10 – 20	Dirty		
10 – 20	(a lot of waste on the shore)		
	Extremely dirty		
20+	(most of the shore is covered		
	with plastic debris)		



## Lab's analysis



Lab's analysis

#### WATER

## $H_2O_2$ 30% + Ehanol

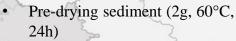
- 50°C
- Orbital stirring
- 24-48 h

Pretreament for eliminating organic matter

#### 1) SAMPLE PRETREATMENT

#### **WET SEDIMENT**

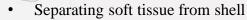
#### NaCl



• Mix sediment with saline solution, allow to settle, collect supernatant (repeat for 3 times)

Pre-treatment with saturated salt solution to float microplastics to the surface

#### **MOLLUSK**



• Weighing the soft tissue

#### $H_2O_2$ 30% + Ethanol

- 50°C
- Orbital stirring
- 1-7 days

Pretreament for eliminating organic matter

#### 2) PURIFICATION WITH OIL

# For greater sample purification: oil can capture all microplastics in the sample

- 1) In the funnel: pre-treated sample + 10 ml olive oil> shake > sediment>discard the watery fraction and mantain the oily (repeat 3 thimes)
- 2) Wash the oily fraction with ethanol and filter

Oily fraction with MP

#### 3) FILTRATION

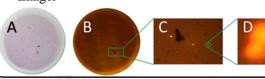
The oily part with microplastics is filtered

Filter: Whatman glass fiber grade GF/F (0,7 µm)

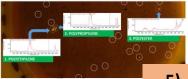
#### 4) QUANTIFICATION

## Nile red staining makes MP glow under UV light

- 10µm/ml in acetone, 30 minutes
- UV Stereomicroscope with orange filter
- ImageJ



## \* Micro-FTIR iN10MX





5) IDENTIFICATIO

EU part of cooperation area

\* Non-EU part of cooperation area

Maes. T., Jessop, R., Wellner, N. et al. A rapid-screening approach to detect and quantify microplastics based on fluorescent tagging



IPA ADRION



**ADRIPLAST** 



## Lab's analysis - UAV





geomorphology, vegetation and marine debris along a sandy spit

Corinne Corbau a,b,c,\*, Joana Buoninsegni a, Elisabetta Olivo a, Carmela Vaccaro a, William Nardin b, Umberto Simeoni c

a University of Ferrara, Ferrara, Italy

b HPL - UMCES, Cambridge, MD, USA

c CURSA, Roma, Italy

## Protocol for quali-quantitative analysis of microplastics in biota



#### 4 STEP



### Delivery and storage of the sample

analyzed within 24 ours after collection or stored in freezer at -20° +/- 5°C





50 organisms -> dissection soft tissue + Hydrogen Peroxide at 30% (20ml / each gram of soft tissue) -> incubation at 55-60° for 4-7 days

#### **Filtration**



Cover the Petri dishes with the appropriate lid and leave the filters to dry at room temperature for at least 24 hours before reading with the stereomicroscope.

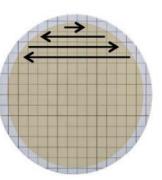


Reading with the stereomicroscope











## MARINE LITTER SURVEYS ON BOCCASETTE BEACH (ROVIGO, ITALY)

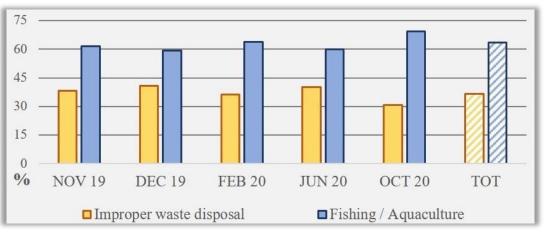
Collected items • 5.578 - 96% artificial polymer materials

Items Density • 0.35 items/m² (± 0.13 SD)

Clean Coast Index

Moderately clean (Nov/Dec-19; Oct-20)

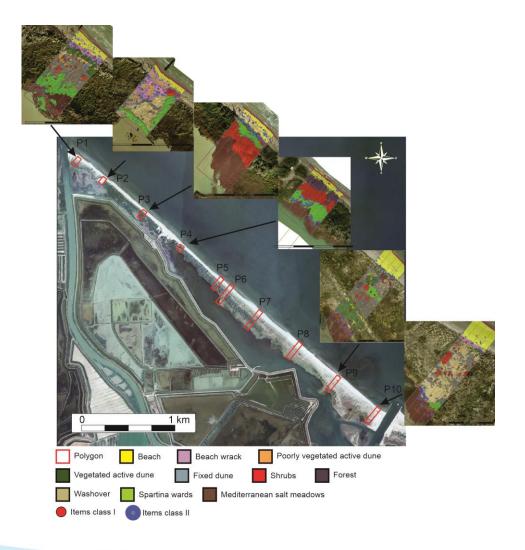
Clean (Feb/Jun-20)



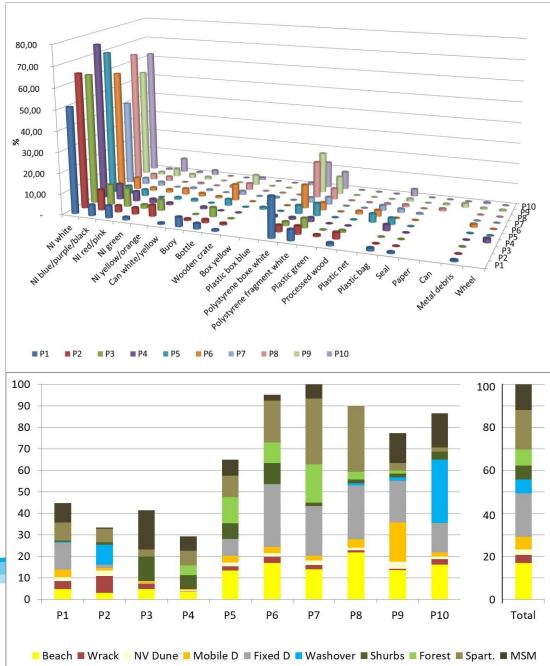
Date	Event notes
Nov 2019	Beginning of 1st sea-storm, high-water and flooding of Po
Dec 2019	After 1st sea-storm; wane high-water and flooding of Po
Feb 2020	After 2 <sup>nd</sup> sea-storm
Jun 2020	Restart of fishing activity and tourism after Covid-19
Oct 2020	After tourist season

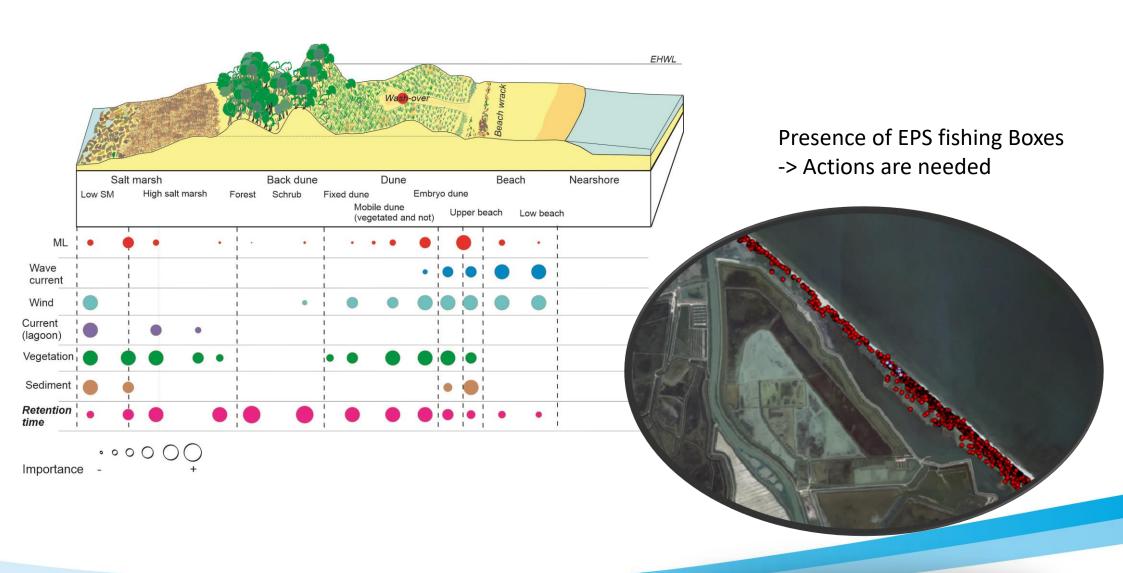
Fishing and Aquaculture > Improper Waste Disposal









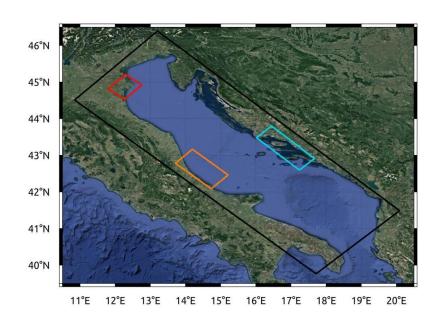


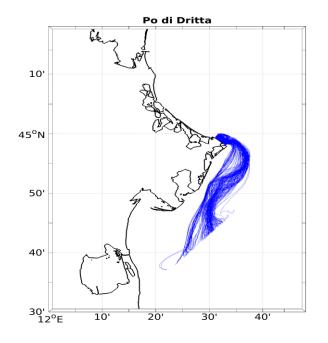


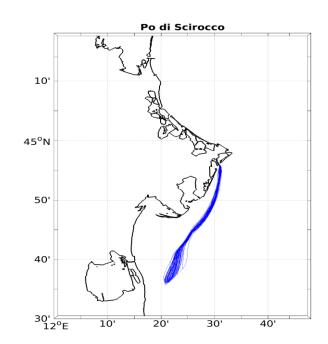


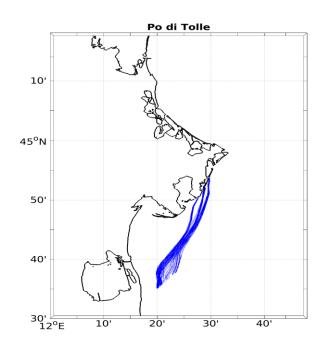
For each site a **4 years long simulation** have been carried out. The analysis of the modeling results has given the possibility to identify the percentage of **potential stranding for fluvial micro-plastics** for each pilot site and to calculate **impact hazard maps for the coasts** both on a daily and climatological basis (yearly, seasonal and monthly).

On following the transport model has been integrated with an **Early Warning System platform.** 



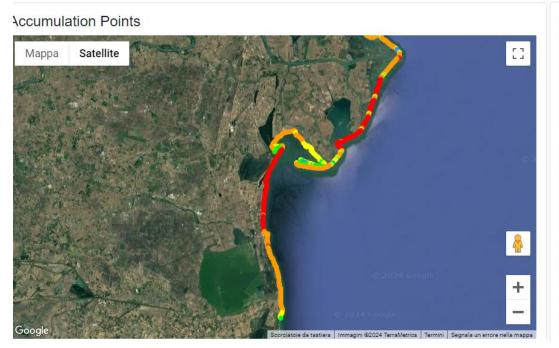


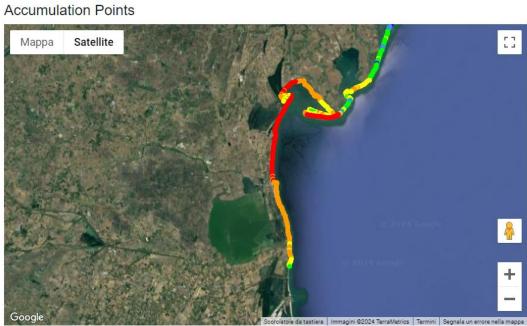






#### Winter – all fluvial contributions





MIN MAX

Winter – Goro Po river contribution

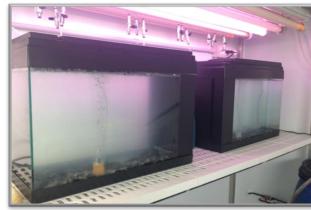






#### **Acclimatization**

- → 7 days in a glass aquarium system (50 l) with filtered artificial seawater aerated at 18 ± 1 °C and salinity 35 ‰.
- → 12 h light-dark regime, no food
- → Gut clearing
- → After 7 days: 20 specimen for blank remaining -> contamination





## **Exposure phase**

- → 3 days
- → A concentration of 2000 MPs/L with a ratio of granules, fragments and fibers of 1: 1: 8 (same proportion found in marine environment according to Su et al., 2016)
- → constant salinity of 35‰, T° of 18 °C, and 12 h light-dark regime
- → Water was renewed, Daily feed, monitoring of the mortality





#### **Purification phase**

- → 20 mussels for control
- → Two different purification times were applied:
  - a "microbiological" purification lasting 2 days;
  - a "experimental" purification lasting 7 days.





## MPs' qualitative and quantitative analysis

For each sampling time

- control group
- 0 time group
- 2 time group7 time group

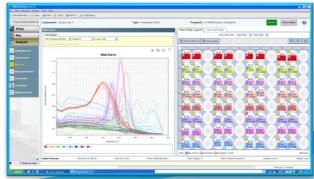
20 organisms were taken and MPs' qualitative and quantitative analysis were performed

#### Gene expression analysis

For each sampling time

- control group
- 0 time group
- 2 time group7 time group
- ✓ 5 organisms were taken and from each separatly gills and digestive gland were taken and pools created
- ✓ In these pools bio-molecular (qPCR real-time) investigations were performed in order to evaluate a set of target genes related



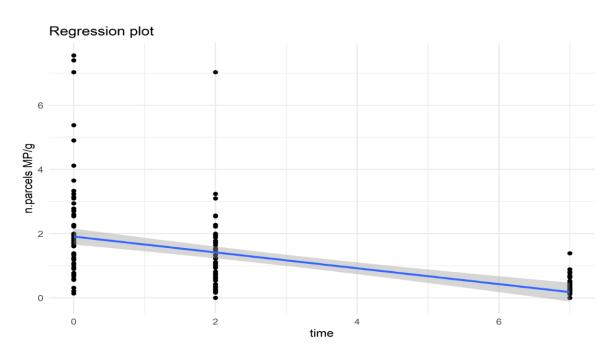






#### Results

The replicates, conducted under the same experimental conditions, did not show statistically significant variability, therefore the experimental error did not interfere with the results obtained.



From the regression plot it is possible to highlight that there is a **statistically significant decrease** (p value 2.5E-14) in the presence of the number of microplastic particles found per gram of soft tissue of the analyzed mussels, in fact we have as average values in the 3 experimental groups:

- Group T0: 2.17 MPs/g;
- Group T2: 0.49 MPs/g;
- Group T7: 0.27 MPs/g.

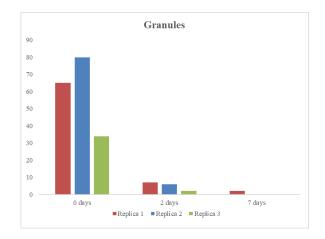
This decrease can be seen already after 2 days of purification and is even more evident after 7 days of purification.

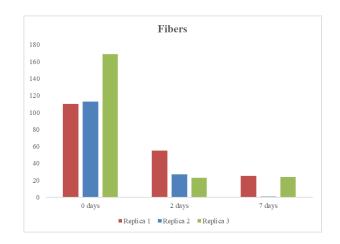
Instead, no statistically significant differences emerged between the T0 and T2 groups indicating that the two-days depuration time may not have been long enough to completely eliminate the microplastic particles or that MP could have been translocated to other tissues, or even to the circulatory system (*Birnstiel et al., 2019; von Moos et al., 2012; Browne et al., 2008*).

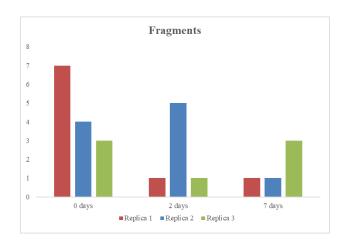




#### Results







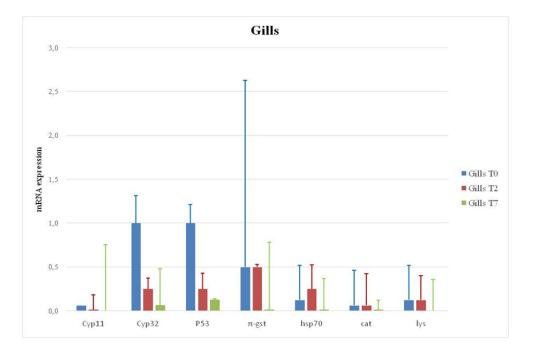
Decrease of the number of each type of microplastic in the three experimental replicas.

More efficient elimination of microplastics : granules -> filaments -> fragments.



## IZSAM CCAPORAL TERAMO

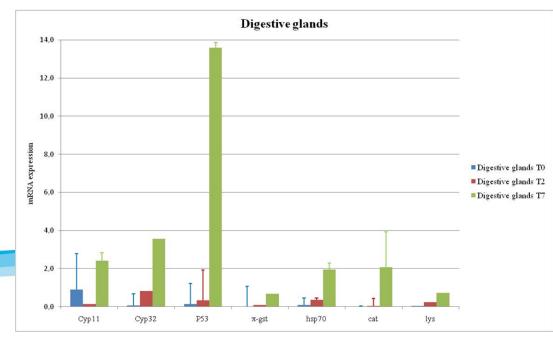
## Gene expression analysis



#### Gene expression data showed that

- In both tested organs we observed differences in contaminated mussels' group or purificated group (T7 group) with respect to the control group.
- In particular, the microplastics altered the expression of genes associated with biotransformation, DNA damage and cell-tissue repair genes

Therefore, the results of the present work confirm that cyp32,  $\pi$ -gst and P53 are transcript biomarker candidates in mussel, in accordance with Lacroix et al. 2014;







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(This article belongs to the Topic Livestock and Microplastics)

**Browse Figures** 

Exposure of *Mytilus galloprovincialis* to Microplastics: Accumulation, Depuration and Evaluation of the Expression Levels of a Selection of Molecular Biomarkers

by Federica Pizzurro 1 2 3. Eliana Nerone 1,\* 3. Massimo Ancora 1 3 3. Marco Di Domenico 1 3 3. Luana Fiorella Mincarelli 1 3. Cesare Cammá 1 3 3. Romolo Salini 1 3 3. Ludovica Di Renzo 1 3 3. Federica Di Giacinto 1 3 3. Corinne Corbau 2 3 3. Itana Bokan 3 3. Nicola Ferri 1 3 3 and Sara Recchi 1 3 1. Istituto Zooprofilattico Sperimentale dell'Abruzzo e Molise (IZSAM), 64100 Teramo, Italy

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**Versions Notes** 

